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Background Information

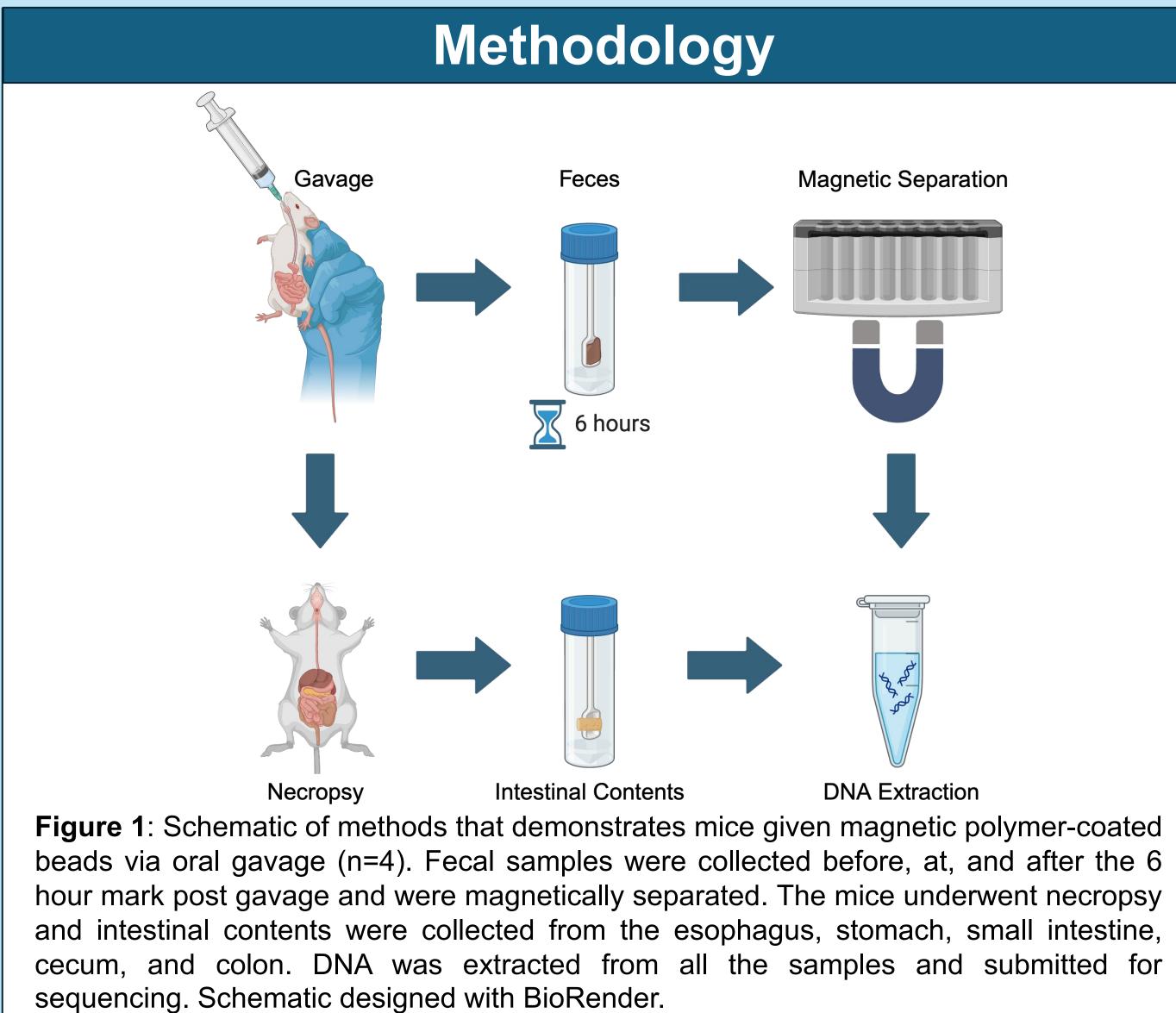
- The microbiome varies between the upper gastrointestinal tract (GIT) and the lower GIT
- Our current methods of measuring the intestinal microbiome rely on fecal testing and thus heavily reflect the lower GIT
- Research has shown that the number of bacteria in the colon is higher than the number of bacteria in the small intestine
- The pH varies along the different segments of the GIT (see Table 1)

Table 1: The pH values of various segments of the mammal GIT

Section of Gastrointestinal Tract	рН
Stomach	1.5-3.5
Duodenum	5.5-6.0
Jejunum	6.0-7.4
lleum	7.4

Objective and Significance

- The objective of this study is to use a novel, proprietary magnetic polymer-coated bead to capture bacteria in the small intestine in a live mouse model
- The significance of this study is to discover the role of the small intestinal microbiome in various disease states, including dysbiosis, intestinal diseases, and even epigenetic factors

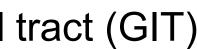


Novel pH-Sensitive Polymer-Grafted Magnetic Microparticles for In Vivo Sampling of the Murine Small Intestine Microbiome

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Histidine Polymer Beads



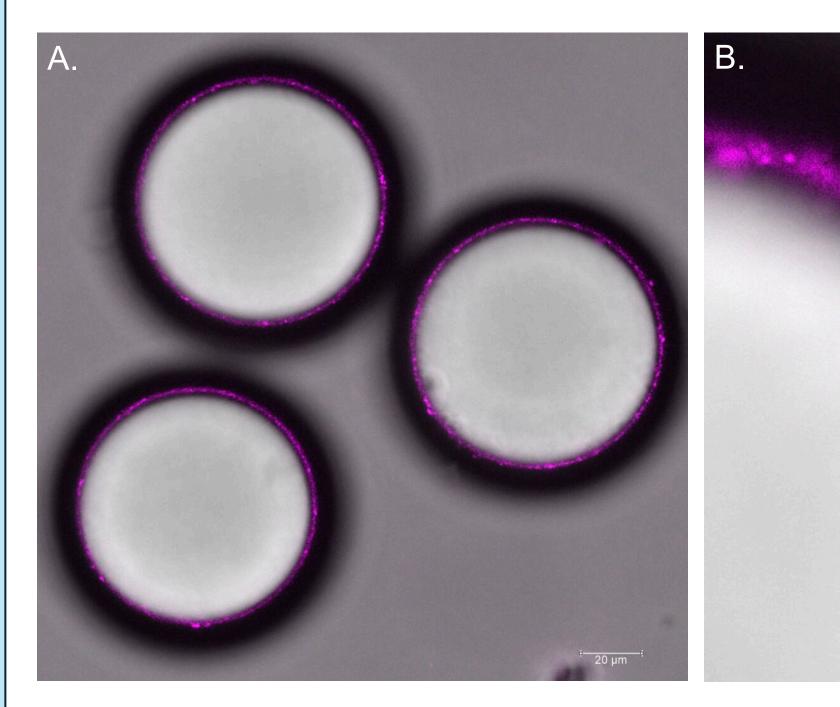


Figure 2: Images of the magnetic polymer-coated beads showing captured bacteria. WGA was used to stain bacteria (magenta). (A) At 100X with a 20 um scale bar (B) Zoomed in using confocal microscopy with a 5 um scale bar

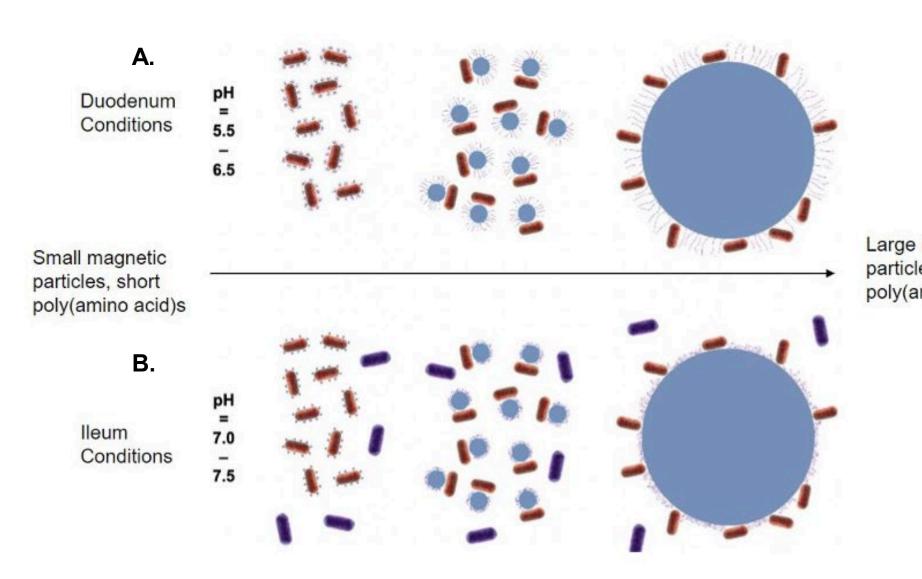
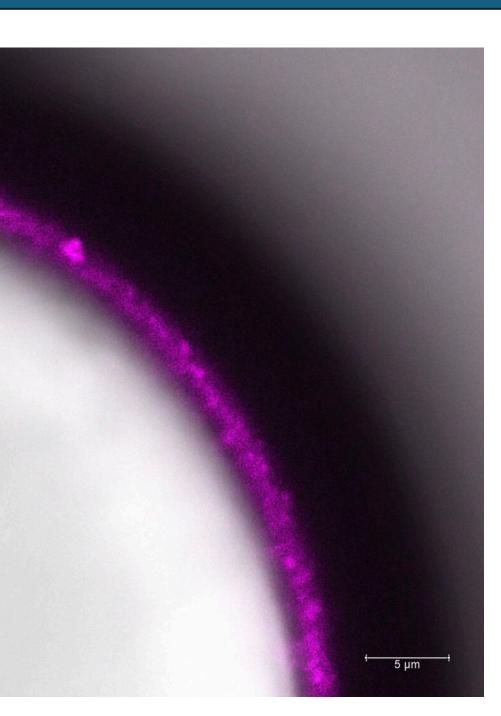


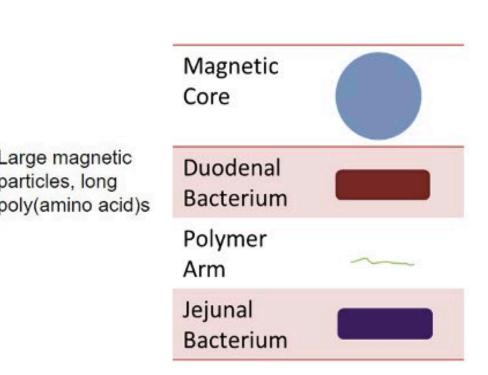
Figure 3: Depiction of the magnetic polymer-coated beads capturing bacteria. The histidine polymer arms are attached to a magnetic bead central core in order to be isolated using a magnet. A small magnetic bead is depicted in the middle segment of the image and a large magnetic bead is depicted in the right side of the image. (A) In the acidic conditions of the duodenum (pH = 5.5-6.5), the histidine polymer arms are in the reduced conformation and contain two positively charged nitrogen groups. In this state, the polymer arms will allow bacteria in the duodenum to attach to them. (B) In the neutral conditions of the lleum (pH = 7.0-7.5), the histidine polymer arms are in the oxidized conformation and contain two neutrally charged nitrogen groups. In this state, the polymer arms hold onto the bacteria that were attached to them from the duodenum, while excluding bacteria that is native to the jejunum and ileum.

Acknowledgements

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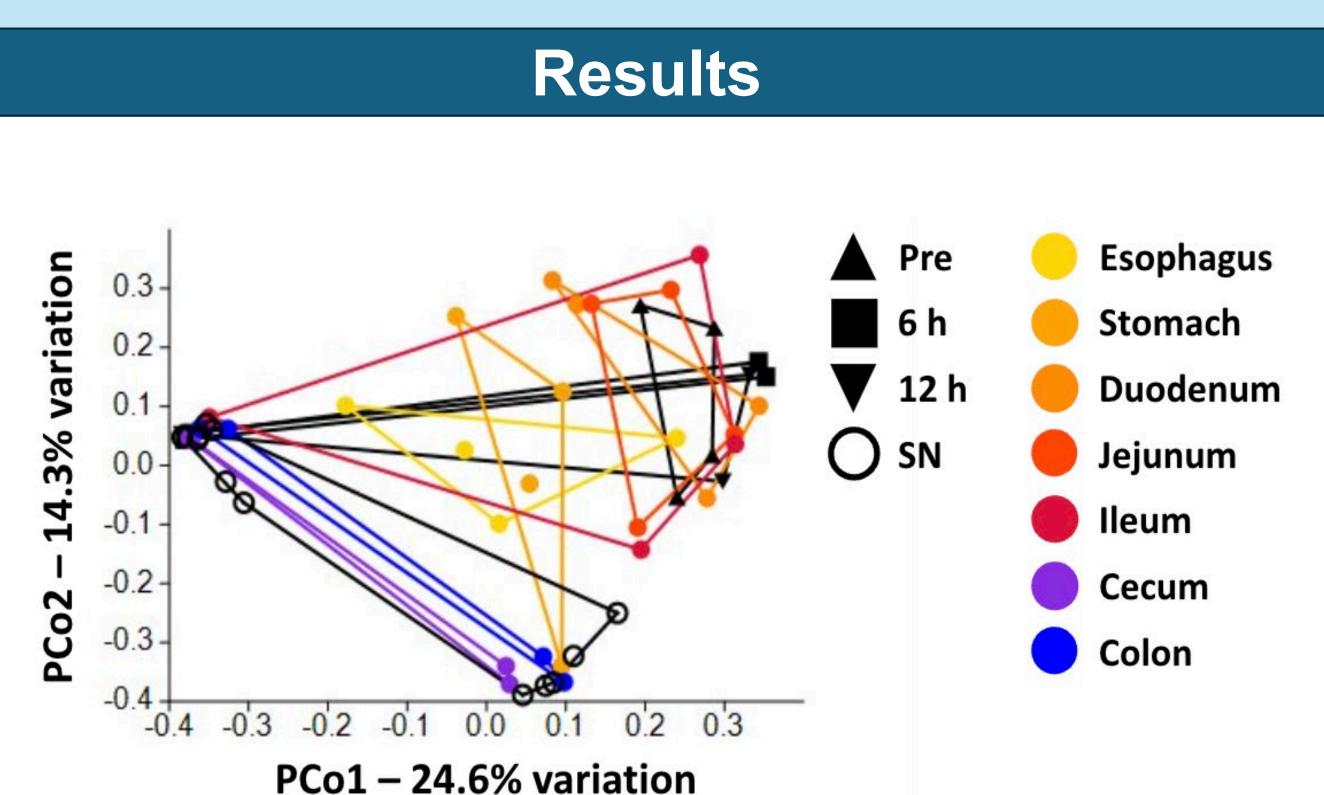


Figure 4: Principal coordinate analysis (PCoA) plot showing Jaccard dissimilarities between capture beads isolated from feces prior to 6 hour (Pre), and at 6 and 12 hours post-administration, the supernatant (SN) from which the beads were separated, and samples collected from different regions of the gastrointestinal tract post-mortem.

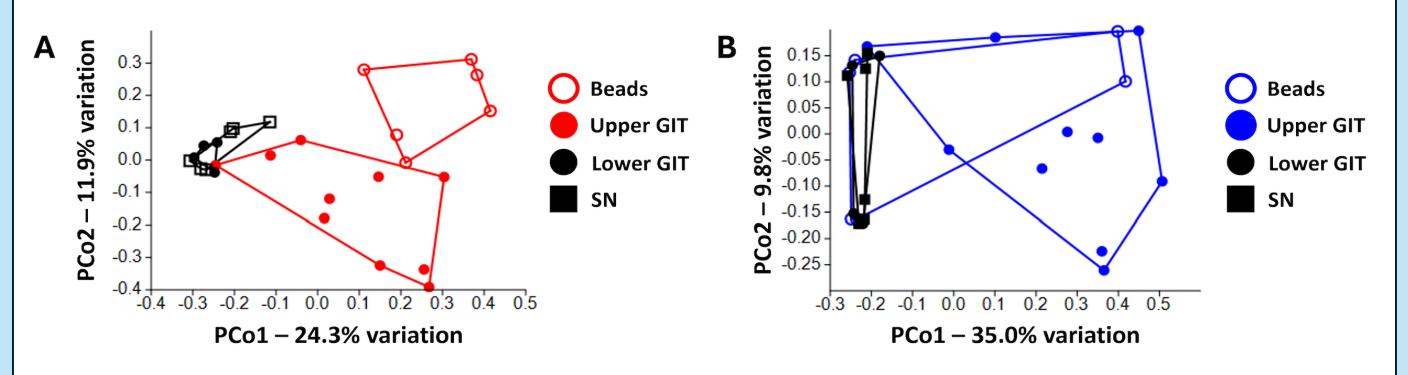


Figure 5: Principal coordinate analysis (PCoA) plot showing Jaccard dissimilarities between capture beads isolated from GM^{low} (A) or GM^{high} (B) feces at 5.5 to 6.5 hours post-administration, the supernatant (SN) from which the beads were separated, and samples collected from upper and lower gastrointestinal tract (GIT) postmortem.

Conclusions

The pH-sensitive polymer grafted particles are readily passed through the GIT These beads can be magnetically isolated from feces The magnetic polymer-coated beads are capturing bacteria from the small intestine, while excluding bacteria from the large intestine ✤Beads isolated from both GM^{low} and GM^{high} mice show binding of small intestinal bacteria

Future Goals

- the mouse GIT
- of the polymer arms at the targeted pH levels
- bacterial contamination



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Measure the transit time of the magnetic polymer-coated beads through

Create an experiment testing the accuracy of the conformation change Analyze the inoculated magnetic polymer-coated beads for any level of